PROTOCOL FOR ANIMAL USE AND CARE
Handwritten forms are not accepted

CRPRC

Investigator
Last Name: [ ]
First: [ ]
Middle: [ ]
email: [ ]
Department: [ ]
Phone / Fax: [ ]
After hrs. #: [ ]

Contact
Last Name: [ ]
First: [ ]
Middle: [ ]
e-mail: [ ]
Department: [ ]
Phone: [ ]
After hrs. #: [ ]

Species (common names): [ ]
Number: [ ]
Source: [ ]

Rhesus macaque 120 CRPRC

Project Title: [ ]
SHIV Immunization to Prevent Vaginal Transmission of SIV

Overnight housing location: [ ]
CRPRC [ ] Day use only:

Animals will be maintained by: [ ]
Vivarium [ ] Investigator
(If investigator maintained, attach husbandry SOP’s.)

Procedures: Provide a one or two sentence layman’s description of the procedures employed on the animals in this project. This information will help the animal care staff understand any conditions they may encounter while caring for your animals.

As part of developing an HIV vaccine, animals will be immunized. Blood, secretions, skin and lymph node biopsies will be collected at regular intervals to assess immune responses and degree of protection from SIV challenge.

Special Husbandry Requirements: Describe any special requirements your animals have with respect to food, water, temperature, humidity, light cycles, caging type, bedding, or any other conditions of husbandry.

None.

Other instructions for animal care staff: (check applicable entries)
Sick Animals: [ ] Call Investigator [ ] Call Investigator [ ] Call Investigator
[ X ] Clinician to treat [ ] Save for Investigator [ X ] OK to use pesticides
[ ] Terminate [ ] Bag for disposal [ ] No Pesticides in animal area
[ ] Necropsy [ X ] Necropsy

Dead Animals: [ X ] Yes [ ] No

Pest Control: [ ] Call Investigator
[ X ] OK to use pesticides
[ ] No Pesticides in animal area

Hazardous Materials (only if in the animal room):
Infectious Agents? [ X ] Yes [ ] No
Agent(s): SIV, SHIV
Radioisotopes? [ ] Yes [ X ] No
Agent(s): 
Chemical Carcinogens? [ ] Yes [ X ] No
Agent(s): 
Toxic Chemicals? [ ] Yes [ X ] No
Agent(s): 
Summary of Procedures:

a) Briefly describe the overall intent of the study. Include in your description a statement of your hypothesis, the objectives and significance of the study. Your target audience is a faculty member from a discipline unrelated to yours. Do not use jargon.

As part of developing an HIV vaccine, animals will be immunized with non-pathogenic SHIV. Blood, secretions, skin and lymph node biopsies will be collected at regular intervals to assess immune responses to the SHIV immunization. Six to twelve months post-immunization the animals will be challenged with pathogenic SIV and monitored for evidence of SIV infection.

b) Procedures employed in this project:

Please check the appropriate boxes if any of these procedures will be employed in your project:

- [ ] Monoclonal Antibody Production **
- [ ] Polyclonal Antibody Production **
- [ ] LD 50 or ID50 studies.
- [X] catheters, blood collection, intubation
- [ ] Prolonged restraint. (8 hrs+)
- [X] Fasting prior to a procedure.

- [ ] Food or water restriction
- [ ] Non-recovery surgical procedures
- [X] Induced illness, intoxication, or disease
- [ ] Survival surgical procedures
- [ ] Multiple survival surgery
- [ ] Behavioral modification.
- [ ] Aversive conditioning.

** If this protocol only describes antibody production, you may use the attached antibody production page in lieu of completing section c below.
c) Describe the use of animals in your project in detail, with special reference to any of procedures checked above. Include any physical, chemical or biological agents that may be administered. List each study group, and describe all the specific procedures that will be performed on each animal in each study group. Use terminology that will be understood by individuals outside your field of expertise. (Note: This cell will expand to whatever length you require. You may make this section as long as you wish, but try to be concise. Some projects may require one or two pages.)

All the animals in every study group will be treated in a similar fashion up to the time of SIV challenge. The animals will be immunized (either intravenously or intranasally), immune responses will be characterized, and then some of the animals will be challenged with SIV. All procedures (collection of blood, secretions, lymph node biopsies and skin biopsies) will be under telazol anesthesia. All animals will be fasted 12 hours prior to ketamine anesthesia. Following a period of 6-12 months, six immunized animals will be necropsied from each group to obtain mucosal and lymphoid tissues for immune response analysis. The other immunized animals will be challenged with SIV. One week after challenge six immunized animals from each group will be necropsied to obtain mucosal and lymphoid tissues for immune response analysis. Two weeks after challenge six immunized animals from each group will be necropsied to obtain mucosal and lymphoid tissues for immune response analysis. We anticipate the last six monkeys from each group to be culled approximately six months post-challenge. We have shown in preliminary studies that SHIV immunized monkeys challenged at six months after immunization are not protected from SIV challenge, while animals challenged at 12 months post-immunization are protected. Thus, we will attempt to determine the nature of the difference in anti-SIV immunity in the two groups. Finally, vaccines are generally used to prevent disease and not infection. Thus, after the SIV challenge, we will monitor six animals in each group until the clinical onset of disease. When an animal develops clinical SIV disease, it will be culled. However, animals may be infected, but apparently healthy, for several years after SIV inoculation and thus animals initiated in one year of the study will be maintained and sampled as described below for several years.

**Group A- Intranasal SHIV Immunization, SIV Challenge 6 Months Post-Immunization** (24 monkeys)

Adult female rhesus macaques will be inoculated with 1 ml SHIV 89.6 intranasally twice a day (AM and PM, approximately 6 hours apart) for two days. Blood samples (not to exceed 12 ml/kg/month) will be obtained at weekly intervals for the first four weeks and every month thereafter. At the time of blood collection, vaginal and rectal secretions will be obtained by lavage with PBS and a cytobrush will be used to atraumatically sample cells in the cervix of the animals. Lymph node biopsies (single lymph node/biopsy) and a 3 mm. diameter skin biopsy will be obtained at 60 day intervals beginning the day of inoculation (day 0). Six months later, the immunized monkeys will be challenged intravaginally with SIVmac239. Six immunized monkeys will be necropsied on the day of challenge. One week after challenge six immunized monkeys will be necropsied. Two weeks after challenge six immunized monkeys will be necropsied. The remaining six animals will be on the same sampling schedule as after the SHIV immunization.

**Group B- Intranasal SHIV Immunization, SIV Challenge 12 Mos. Post-Immunization** (24 monkeys)

Adult female rhesus macaques will be inoculated with 1 ml SHIV 89.6 intranasally twice a day (AM and PM, approximately 6 hours apart) for two days. Blood samples (not to exceed 12 ml/kg/month) will be obtained at weekly intervals for the first four weeks and every month thereafter. At the time of blood collection, vaginal and rectal secretions will be obtained by lavage with PBS and a cytobrush will be used to atraumatically sample cells in the cervix of the animals. Lymph node biopsies (single lymph node/biopsy) and a 3 mm. diameter skin biopsy will be obtained at 60 day intervals beginning the day of inoculation (day 0). Twelve months later, the immunized monkeys will be challenged intravaginally with SIVmac239. Six immunized monkeys will be necropsied on the day of challenge. One week after challenge six immunized monkeys will be necropsied. Two weeks after challenge another six monkeys will be necropsied. The remaining six animals...
will be on the same sampling schedule as after the SHIV immunization.

**Group C— Intravenous SHIV Immunization, SIV Challenge 6 Months Post-Immunization (24 monkeys)**

Adult female rhesus macaques will be inoculated with 1 ml SHIV 89.6 intravenously twice a day (AM and PM, approximately 6 hours apart) for two days. Blood samples (not to exceed 12 ml/kg/month) will be obtained at weekly intervals for the first four weeks and every month thereafter. At the time of blood collection, vaginal and rectal secretions will be obtained by lavage with PBS and a cytobrush will be used to atraumatically sample cells in the cervix of the animals. Lymph node biopsies (single lymph node/biopsy) and a 3 mm. diameter skin biopsy will be obtained at 60 day intervals beginning the day of inoculation (day 0). Six months later, the immunized monkeys will be challenged intravaginally with SIVmac239. Six immunized monkeys will be necropsied on the day of challenge. One week after challenge six immunized monkeys will be necropsied. Two weeks after challenge six immunized monkeys will be necropsied. The remaining animals will be on the same sampling schedule as after the SHIV immunization.

**Group D— Intravenous SHIV Immunization, SIV Challenge 12 Mos. Post-Immunization (24 monkeys)**

Adult female rhesus macaques will be inoculated with 1 ml SHIV 89.6 intravenously twice a day (AM and PM, approximately 6 hours apart) for two days. Blood samples (not to exceed 12 ml/kg/month) will be obtained at weekly intervals for the first four weeks and every month thereafter. At the time of blood collection, vaginal and rectal secretions will be obtained by lavage with PBS and a cytobrush will be used to atraumatically sample cells in the cervix of the animals. Lymph node biopsies (single lymph node/biopsy) and a 3 mm. diameter skin biopsy will be obtained at 60 day intervals beginning the day of inoculation (day 0). Twelve months later, the immunized monkeys will be challenged intravaginally with SIVmac239. Six immunized monkeys will be necropsied on the day of challenge. One week after challenge six immunized monkeys will be necropsied. Two weeks after challenge another six monkeys will be necropsied. The remaining animals will be on the same sampling schedule as after the SHIV immunization.

**Group E— Control Animals (24 monkeys)**

Adult female rhesus macaques will be challenged intravaginally with SIVmac239. These animals are not immunized with SHIV and are necessary to demonstrate the infectivity of the challenge virus stock. Blood samples (10 ml or not to exceed CRPRC guidelines) will be obtained at weekly intervals for the first four weeks and every month thereafter. At the time of blood collection, vaginal and rectal secretions will be obtained by lavage with PBS and a cytobrush will be used to atraumatically sample cells in the cervix of the animals. Lymph node biopsies (single lymph node/biopsy) and a 3 mm. Diameter skin biopsy will be obtained at 60 day intervals beginning the day of inoculation (day 0). Six monkeys will be necropsied on the day of challenge. One week after challenge six monkeys will be necropsied. Two weeks after challenge another six monkeys will be necropsied. The remaining six animals will be on the same sampling schedule as the immunized monkeys and will be culled before the development of clinical SAIDS.

d) Study Groups and Numbers: Define, in the form of a table, the numbers of animals to be used in each experimental group described above. The table may be presented on a separate page as an attachment to this protocol if you prefer. The Normal format should be three columns: Study Group, Procedure, Number of animals. The number of rows should follow from the number of study groups; you may add as many rows as you require. The chart must fully account for the number of animals you intend to use under this protocol. Assign each group to an invasiveness category according to the chart below.

<table>
<thead>
<tr>
<th>Group</th>
<th>Procedures / Drugs</th>
<th>Number of Animals</th>
<th>Category</th>
</tr>
</thead>
<tbody>
<tr>
<td>A</td>
<td>SHIV intranasal immunization, SIV challenge 6 mos.</td>
<td>24</td>
<td>4</td>
</tr>
<tr>
<td></td>
<td>SHIV intranasal immunization, SIV challenge 12 mos.</td>
<td>24</td>
<td>4</td>
</tr>
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<td>--------------------------------------------------</td>
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</tr>
<tr>
<td>C</td>
<td>SHIV intravenous immunization, SIV challenge 6 months</td>
<td>24</td>
<td>4</td>
</tr>
<tr>
<td>D</td>
<td>SHIV intravenous immunization, SIV challenge 12 months</td>
<td>24</td>
<td>4</td>
</tr>
<tr>
<td>E</td>
<td>SIV challenge</td>
<td>24</td>
<td>4</td>
</tr>
</tbody>
</table>

**Categories of invasiveness**

<table>
<thead>
<tr>
<th>Category</th>
<th>Description</th>
</tr>
</thead>
</table>
| 1        | Little or no discomfort or stress  
Examples: domestic flocks or herds being maintained in simulated or actual commercial production management systems; the short-term and skillful restraint of animals for purposes of observation or physical examination; blood sampling; injection of material in amounts that will not cause adverse reactions by the following routes: intravenous, subcutaneous, intramuscular, intraperitoneal, or oral. |
| 2        | Minor stress or pain of short duration  
Examples: cannulation or catheterization of blood vessels or body cavities under anesthesia; minor surgical procedures under anesthesia, such as biopsies or laparoscopy; short periods of restraint beyond that required for simple observation or examination, but consistent with minimal distress |
| 3        | Moderate to severe distress  
Examples: major surgical procedures conducted under general anesthesia, with subsequent recovery; prolonged (several hours or more) periods of physical restraint; induction of behavioral stresses such as maternal deprivation |
| 4        | Severe pain near, at or above the pain tolerance threshold  
Examples: exposure to noxious stimuli or agents whose effects are unknown; exposure to drugs, chemicals, or infectious agents at levels that markedly impair physiological systems and which cause death, severe pain, or extreme distress: Surgical experiments which have a high degree of invasiveness. |

Further descriptions of these categories are included in the instructions following this document.

e) Rationale for species and numbers: How did you determine that 1) the species choice was appropriate and 2) the number of animals in each study groups was the minimum number necessary to achieve sound scientific results?

Rhesus macaques are the only animal model in which reliable data regarding vaccine efficacy for protection from vaginal SIV challenge can be obtained. We have settled on group sizes of 6 for all studies. Thus each group of 24 immunized monkeys will be challenged, six culled the day of challenge, six one week post-challenge and six two weeks post-challenge, with six remaining until necropsy approximately 6 months later. There are 24 control monkeys, six to be necropsied one week post-challenge, six two weeks post-challenge, and we anticipate the rest being culled approximately 6 months post-challenge. Based on a student T test, these are the smallest groups that can be used to detect a significant difference in the outcome of the immunized and control animals. Note that because of the labor-intensive nature of the analysis, it is necessary to challenge groups A-D on different calendar days. Thus, six animals from group E will be used each time animals from the other four groups are challenged.

f) Surgery: If the project involves survival surgery, where will the surgery be conducted?

Building:  
Room:  
Who will be the surgeon:  

g) Anesthetics, Analgesics, Tranquilizers, Neuromuscular blocking agents:
Post procedural analgesics should be given whenever there is possibility of pain or discomfort that is more than slight or momentary. If postoperative analgesics are not to be given, justify the practice under part (i) below.

Provide the following information about any of these drugs that you intend to use in this project.

<table>
<thead>
<tr>
<th>Species</th>
<th>Drug</th>
<th>Dose (mg/kg)</th>
<th>Route</th>
<th>When and how often will it be given?</th>
</tr>
</thead>
<tbody>
<tr>
<td>rhesus</td>
<td>Telazol</td>
<td>6-8 mg/kg</td>
<td>IM</td>
<td>Before all procedures</td>
</tr>
<tr>
<td>rhesus</td>
<td>Oxymorphone</td>
<td>1 mg/kg</td>
<td>IM</td>
<td>As needed in the judgement of CRPRC vets</td>
</tr>
</tbody>
</table>

h) Neuromuscular blocking agents can conceal inadequate anesthesia and therefore require special justification. If you are using a neuromuscular blocking agent, please complete the following:

Why do you need to use a neuromuscular blocking agent?

N/A

What physiologic parameters are monitored during the procedure to assess adequacy of anesthesia?

Under what circumstances will incremental doses of anesthetics-analgesics be administered?

i) Adverse effects:

Describe any potential adverse effects of the experiment on the animals (such as pain, discomfort; reduced growth, fever, anemia, neurological deficits; behavioral abnormalities or other clinical symptoms of acute or chronic distress or nutritional deficiency)

Any injection or venipuncture has the potential to cause minor pain or discomfort, but the animals are immobilized for the procedure and should not experience pain.

SIV infection of rhesus macaques results in a fatal immunodeficiency and wasting syndrome. The animals will be euthanized before, or when, they experience 3 of the following: weight loss >15% in two weeks or >30% in 3 months; persistent hypothermia <96°F even with heat supplementation; leukopenia (total WBC<3,000); lymphopenia (lymphocytes <800); anemia (hemoglobin <10); dehydration >10%; nonresponsive to therapy for opportunistic infections; persistent anorexia (>3 days); animal significantly obtunded. These criteria are based on CRPRC guidelines. In addition, the lymph node biopsies will result in some post-procedure pain.

How will the signs listed above be ameliorated or alleviated? If signs are not to be alleviated or ameliorated by means of post-operative analgesics or other means, explain why this is necessary.

All possible efforts will be made to minimize animal pain and discomfort. Analgesics have no effect on the proposed studies and they will be administered at the discretion of the CRPRC veterinary staff. The SIV infected animals will be euthanized prior to or at the time they develop clinical signs of AIDS. The decision to euthanize will be based on the judgement of the CRPRC veterinarians.

Note: if any unanticipated adverse effects not described above do occur during the course of the study, a complete description of those effects and the steps taken to mitigate them must be submitted to the committee as an amendment to this protocol.
Is death an endpoint in your experimental procedure?  [ ] Yes  [X] No

(Note: "Death as an endpoint" refers to acute toxicity testing, assessment of virulence of pathogens, neutralization tests for toxins, and other studies in which animals are not euthanized, but die as a direct result of the experimental manipulation). If death is an endpoint, explain why it is not possible to euthanize the animals at an earlier point in the study. If you can euthanize the animals at an earlier point, describe the clinical signs which will dictate that an animal will be euthanized.

j) Literature search for alternatives and unnecessary duplication:

This section is specifically required by Federal law. You are required to conduct a literature search to determine that either 1) there are no alternative methodologies by which to conduct this study, or 2) there are alternative methodologies, but these are not appropriate for your particular study. "Alternative methodologies" refers to reduction, replacement, and refinement (the three R's) of animal use, not just animal replacement. You must also show that the study is not unnecessarily duplicative of other studies.

What was the date on which you conducted this search?  4/17/01

List the databases searched or other sources consulted (there should be more than one). Include the years covered by the search.

<table>
<thead>
<tr>
<th>Database Name</th>
<th>Years Covered</th>
<th>Keywords / Search Strategy</th>
</tr>
</thead>
<tbody>
<tr>
<td>Medline</td>
<td>unlimited</td>
<td>SIV, SHIV, HIV, vaginal transmission, vaccines, protection</td>
</tr>
<tr>
<td>AIDSline</td>
<td>unlimited</td>
<td>SIV, SHIV, HIV, vaginal transmission, vaccines, protection</td>
</tr>
<tr>
<td>Current Contents</td>
<td>unlimited</td>
<td>SIV, SHIV, HIV, vaginal transmission, vaccines, protection</td>
</tr>
</tbody>
</table>

What were your findings with respect to alternative methodologies?

The only available animal model systems of HIV heterosexual transmission are the SIV/rhesus macaque, HIV/chimpanzee and FIV/cat. No other animal models are satisfactory for assessing the ability of vaccines to prevent vaginal HIV transmission. In-vitro systems are unsatisfactory for these studies.

Has this study been previously conducted?  [ ] Yes  [X] No

If the study has been conducted previously, explain why it is scientifically necessary to replicate the experiment.

k) Disposition of animals: At what point in the study, if any, will the animals be euthanized?

Animals will be euthanized for obtaining samples as noted in section C. In addition, the SIV infected animals will be euthanized prior to, or at the time they develop clinical signs of AIDS. (see 19a above)

l) Methods of euthanasia: Even if your study does not involve killing the animals, you should show a method that you would use in the event of unanticipated injury or illness. If anesthetic overdose is the method, show the agent, dose, and route.

<table>
<thead>
<tr>
<th>Species</th>
<th>Method</th>
<th>Drug</th>
<th>Dose (mg/kg)</th>
<th>route</th>
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<tbody>
<tr>
<td>rhesus</td>
<td>IV</td>
<td>pentobarbital</td>
<td>60 mg/kg</td>
<td>IV</td>
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</table>

m) Surplus animals: What will you do with any animals not euthanized at the conclusion of the project?

All animals will be euthanized at the end of the project.
n) **Project Roster:** Please provide the names of all the individuals who will work with animals on this project. This page will not be made available to the public. Give either the University Employee ID # or a valid UC Davis email address so that we can document training and occupational health compliance for regulatory agencies. Include all investigators, student employees, post-doctoral researchers, staff research associates, post-graduate researchers and laboratory assistants who will actually work with the animals. You don't need to include the staff of the vivarium in which your animals will be housed.

The principal investigator is responsible for keeping this roster current. If any staff is added or subtracted from this project, you must amend the protocol by sending the campus veterinarian a memo describing any changes.

<table>
<thead>
<tr>
<th>Last Name</th>
<th>First Name</th>
<th>Middle Name</th>
<th>UC ID Number or SSN</th>
<th>Email Address</th>
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**Occupational Health Program:**

Supervisors must enroll their employees in the campus Occupational Health Program if the workers are at increased risk of illness or injury (such as allergy, physical injury, or infectious disease) because of their work. Enroll workers by having them complete an "Animal Contact History Form", available from Employee Health Services (phone 752-2330). For further information, visit our web site at [http://clueless.ucdavis.edu/health/](http://clueless.ucdavis.edu/health/) or read the UC Davis Policy & Procedure Manual 290-25.

**Training:**

Supervisors are responsible for insuring that their employees are adequate trained, both in the specifics of their job and in the requirements of the Federal Animal Welfare Act. EH&S offers free, basic wet labs in laboratory animal handling and techniques, and lecture format classes in the requirements of the Animal Welfare Act. To schedule a class for your unit, contact EH&S at 2-2364. Autotutorials are also available on the world wide web at [http://clueless.ucdavis.edu/](http://clueless.ucdavis.edu/).
Assurances for the Humane Care and Use of Vertebrate Animals:

Principal Investigator's Statement:

I have read and agree to abide by the UC Davis Policy and Procedure Manual section 290-30 (Animal Use and Care). This project will be conducted in accordance with the ILAR Guide for the Care and Use of Laboratory Animals, and the UC Davis Animal Welfare Assurance on file with the US Public Health Service. (These documents are available from the Campus Veterinarian and at http://ehs.ucdavis.edu/). I will abide by all Federal, state and local laws and regulations dealing with the use of animals in research.

I will advise the Animal Use and Care Administrative Advisory Committee in writing of any significant changes in the procedures or personnel involved in this project.

___________________________________________  ____________________________________________  ____________________________
Principal Investigator                                         Rank / Title                                         Date

Committee Use Only Below

** Conditions necessary for Committee Approval:


Final Disposition of this protocol:

___________ Approved

___________ Not Approved

___________ Withdrawn by Investigator

Date of Action: _____ / _____ / ______

I verify that the Institutional Animal Care and Use Committee of the University of California, Davis, acted on this protocol as shown above.

___________________________________________  ____________________________
Campus Veterinarian                                         Date
ANIMAL ROOM SAFETY INFORMATION

Complete this form if you will be using biohazards, radioisotopes, carcinogens, or toxic chemicals in the animal room.

Identity of Hazard: SIV, SHIV

Investigator Last Name: ___________________________  Department: ___________________________
First Name: ___________________________  Phone: ___________________________
Email: ___________________________  Fax: ___________________________

Provide a short description of the agent:

SIV and SHIV are primate lentiviruses which can infect human cells and potentially humans.

This agent / material is hazardous for: [ ] Humans only  [ ] Animals only  [X] Humans and Animals
For which Animal Species?

The agent can be spread by: [X] Blood  [ ] Feces/urine  [ ] Saliva/nasal droplets  [ ] Does not leave animal

Describe any human health risk associated with this agent:

No human disease related to these viruses has ever been described. However, there is a potential for these viruses to infect humans.

The precautions checked below apply to this experiment:

[ ] The researcher or his/her technicians are responsible for the feeding and care of these animals.
[X] The following items must be assumed to be contaminated with hazardous material and must be handled only by the researcher or his/her technicians.

[X] Bedding  [ ] Other:

[ ] Cages must be autoclaved before cleaning.
[ ] Label cages and remove label after decontamination.
[ ] Animal carcasses must be labeled and disposed of as follows:

[ ] Incineration  [X] Biohazardous Waste Container
[ ] Bag and Autoclave  [ ] EH&S will pick-up (2-1493).

[ ] All contaminated waste (soiled bedding or other animal waste) must be properly labeled and disposed of as follows

[ ] Incineration  [X] Biohazardous Waste Container
[ ] Bag and Autoclave  [ ] EH&S will pick-up (2-1493).

Personal Protective Equipment Required:

[X] The following personal protective equipment must be worn/used in the room:

[X] Lab Coat/Coveralls  [X] Shoe Covers/Booties
[X] Disposable Gloves  [X] Head Cover
[ ] NIOSH Certified Dust Mask  [ ] Disinfectant footbath
[X] Eye Protection/Face Shield  [ ]
[ ] Fitted Respirator  Type:  [ ] Other:  Describe:

[X] Personal protective equipment must be removed before leaving the room.
[X] Personal protective equipment must be discarded or decontaminated at the end of the project
[X] Hands, arms, and face must be thoroughly washed upon leaving the room.
[ ] Full shower, including washing of hair, must be taken upon leaving the room.
[ ] Decontaminate Room (inform ARS area supervisor when cage and/or room can be returned to general use).

Provide any other information needed to safely work in this room: